Lipase-mediated transformation of D-xylose and D-galactose into less common L-aldoses

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Xylitol and dulcitol have been converted into L-xylose and L-fucose, respectively, by employing lipase-mediated enantioselective deacylation or acylation of their cyclic acetal derivatives as the key step.

As a part of our studies on the enantioselective hydrolysis of various racemic esters catalysed by porcine pancreatic lipase (PPL), the hydrolysis of 1-*O*-acetyl-2,4:3,5-di-*O*-methylidene-DL-xylitol was shown to afford 2,4:3,5-di-*O*-methylidene-L-xylitol of *ca.* 100% enantiomeric purity.¹ In principle, this transformation might be used as a short cut to the 'unnatural' L-xylose from its abundant D-counterpart; however, the difficulty of removing the *O*-methylidene protecting groups was an obstacle to achieving this. On the other hand, PPL-catalysed hydrolysis of the corresponding di-*O*-benzylidene derivative was practically devoid of enantioselectivity.¹

Now we report the preparation of L-xylose (L-1) from xylitol 2 using PPL-catalysed hydrolysis of 1-*O*-acetyl-2,4:3,5-di-*O*-ethylidene-DL-xylitol (rac-3a) as the key step. Acetate rac-3a was obtained from 2 in two conventional steps via racemic alcohol rac-3 (Scheme 1). In contrast with its di-*O*-methylidene analogue, at 45% conversion rac-3a gave not levorotatory, but dextrorotatory alcohol D-3 { $[\alpha]_D^{22}$ +3.6°, (c 1.0, H₂O)}.† Using Pfitzner–Moffatt oxidation, the latter was converted into aldehyde D-4 {mp 164 °C (sublimed), $[\alpha]_D^{22}$ +13.4°, (c 1.0, H₂O)} which was deprotected to afford practically enantiopure D-xylose { $[\alpha]_D^{22}$ +73.9° (15 min) \Rightarrow +18.8° (2 h), (c 1.0, H₂O)}. The enantiomeric purity of intermediate D-3 was confirmed by the ¹⁹F and ¹H NMR spectra of its (S)-MTPA ester.

In order to obtain the required enantiopure alcohol L-3, enzymatic hydrolysis of rac-3a was extended to 55% conversion. Column chromatography on SiO₂ or, better, partitioning of the products between the aqueous phase and CHCl₃ followed by filtration of the concentrated organic layer through a pad of SiO₂ using hexane–AcOEt (2:1, v/v) as the eluent afforded acetate L-3a as a microcrystalline solid {mp 55–56 $^{\circ}\text{C}$ (from petroleum ether, -60 °C), $[\alpha]_{D}^{22}$ +7.30°, (c 1.0, CHCl₃)}. This was saponified to give enantiopure L-3 { $[\alpha]_D^{22}$ -3.5°, (c 1.0, H₂O)}[†] whose ¹H and ¹³C NMR spectra contained only signals attributable to the diequatorial diastereoisomer, while its (S)-MTPA ester displayed no signals attributable to the D-counterpart in its ¹⁹F and ¹H NMR spectra. Oxidation of L-3 resulted in 2,4:3,5-di-O-ethylidene-L-xylose (L-4) with mp 162–164 °C (sublimed) and $[\alpha]_D^{22}$ –13.4°, (c 1.0, H_2O). Lit.,² mp 152–160 °C, $[\alpha]_D^{20}$ –13.2° (H_2O). Acid-catalysed hydrolysis of diacetal L-4 gave the target L-xylose with mp 145-146 °C (from EtOH) and $[\alpha]_D^{22}$ -76.3° (15 min) \rightarrow -18.4° (2 h) (c 1.0, H₂O). Lit.,³ mp 144 °C, $[\alpha]_D^{20}$ -79.3° \rightarrow -18.6° (H₂O).

Allowing for 55% conversion of *rac-3a* at the enzymatic kinetic resolution step, the material yield of L-1 from 2 is about 16% over six steps of the synthesis. This is comparable with earlier syntheses of L-1 from other sugars.^{3,4} Since *meso*-pentaol 2 is obtained directly from D-1, this work represents a formal synthesis of L-1 from D-1 based on controlled asymmetrization of a *meso* precursor ('*meso*-trick').

An alternative PPL-mediated approach from 2 to L-1,

involving the transformation of **2** into 1-*O*-acetyl-2,3:4,5-di-*O*-isopropylidene-DL-xylitol (rac-**5a**) via the corresponding alcohol (rac-**5**)⁵ and enzymatic hydrolysis of rac-**5a** to 35–45% conversion, proved to be inefficient. In this case the e.e. of the resulting alcohol L-**5** {mp 34–35 °C (from hexane, -60 °C), $[\alpha]_D^{22} + 3.25^\circ$ (c 2.0, EtOH)} was only 26–28%. Lit. (for L-**5**),6 mp 33–35 °C (from hexane, -60 °C), $[\alpha]_D^{18} + 12.5^\circ$ (c 2.0, EtOH). When the unconverted fraction of the acetate (i.e., mainly D-**5a**) was again hydrolysed in the presence of PPL to 38% conversion, and residual D-**5a** was saponified, the specimen of alcohol D-**5** thus obtained {mp 33–35 °C (from hexane, -60 °C), $[\alpha]_D^{22} - 3.6^\circ$ (c 2.0, EtOH)} had about 29% e.e.

Another example of lipase-mediated 'meso-trick' strategy in carbohydrate chemistry is represented by the formal synthesis

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 $^{^\}dagger$ Although our specimens of D-3 and L-3 had mp 139–140 °C (from AcOEt), whereas earlier² mp 164–165 °C was reported for L-3, the $[\alpha]_{\rm D}$ values of both specimens almost coincided with those reported in ref. 2, and the NMR spectra of their (S)-MTPA esters were in good agreement with the assigned structures.

of L-fucose **6** from D-galactose, where dulcitol **7** was used as the starting material. At the beginning [Scheme 2(A)], **7** was converted into a mixture of two isomeric diacetonides, where the major component had a symmetric *meso* structure **8** and the minor one was racemic $(rac-9 \equiv 9 + ent-9, 1:1)$. The required *meso* diol **8** was isolated from its mixture with rac-9 either by fractional crystallisation (cf. ref. 7) or by converting this mixture into the corresponding diacetates **8a** and rac-9a, isolating the poorly soluble **8a** by recrystallisation, and saponifying it back to **8**. Originally, it was planned to asymmetrize **8a** by PPL-catalysed hydrolysis. However, even at rather low substrate-to-enzyme ratios (**8a**: PPL = 1:2, w/w) and long exposures (72–120 h) no conversion of **8a** was detected.

As an alternative, controlled acylation of diol **8** using vinyl acetate and the lipase from *Candida rugosa* (= *C. cylindracea*, CCL, Fluka, 2U mg⁻¹) as the catalyst was

C

12
$$\frac{Me}{98\%}$$
 $\frac{Me}{H}$
 $\frac{ix}{48\%, \text{ from } 13 \text{ $via } 14}$
 $\frac{ix}{HO}$
 $\frac{Me}{HO}$
 $\frac{Me}{HO}$
 $\frac{Me}{Me}$
 $\frac{Me}{HO}$
 $\frac{Me}{HO}$
 $\frac{Me}{HO}$
 $\frac{Me}{HO}$
 $\frac{13}{48\%, \text{ from } 13 \text{ $via } 14}$
 $\frac{14}{HO}$
 $\frac{14}{HO}$
 $\frac{14}{HO}$
 $\frac{14}{HO}$
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 $\frac{14}{HO}$
 $\frac{14}{HO}$$$

undertaken [Scheme 2(B)]. At optimal exposures (19–23 h) the yield of levorotatory monoacetate $10~\{\rm mp~71–72~^{\circ}C$ (from Et_2O–hexane), $[\alpha]_D^{22}$ –9.02° $(c~1.0,{\rm CHCl_3})\}$ amounted to 40–43%, the recovery of diol 8 and the yield of diacetate 8a being 47% and 8%, respectively. Longer exposures increased the yield of $10~\rm up$ to 73%, but at the expense of e.e. ($[\alpha]_D^{22}$ –6.4° after 44 h). Mesylation of $10~\rm and$ subsequent treatment of mesylate $11~\rm with$ NaI led to the wax-like iodide $12~\rm (mp~45–47~^{\circ}C)$ which was cleanly hydrogenolysed (with concomitant deacylation) over skeletal Ni in the presence of K_2CO_3 in MeOH to give the known 8 2,3:4,5-di-O-isopropylidene-L-fucitol $13~\rm with$ mp 59–59.5 °C (from hexane) and $[\alpha]_D^{22}$ +11.63° $(c~1.0,{\rm EtOH})$.

Finally, alcohol 13 was oxidised into the corresponding oxo-diketal 14, and the latter was immediately hydrolysed to give the target sugar 6 {mp 138–140 °C (from EtOH), $[\alpha]_D^{22}$ –110° (15 min) \rightarrow –74.7° (4 h) (c 0.95, H₂O)}. Lit., {mp 137–139 °C (from EtOH), $[\alpha]_D$ –75°(24 h) (c 0.95, H₂O)}. Taking into account the content of 8 in the starting mixture of isomeric diols, the yield of L-fucose from 7 was ca. 6% over nine steps. This is comparable with earlier syntheses of 6 from other sugar derivatives. 9.10

Our results confirm the usefulness of lipases in carbohydrate synthesis (*cf.* reviews 11 and 12).

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